

Comparison Between Antibacterial Extract of Gambier (UncariagambirRoxb) and Chlorhexidine 2% to Enterococcus Faecalis

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ABSTRACT

Background: In the case of reinfection after endodontic treatment, *Enterococcus faecalis* is a bacterium that is commonly found in root canals. Irrigation was carried out to eliminate *Enterococcus faecalis* from the root canal. Chlorhexidine 2% irrigating agent was effective in reducing the growth of *Enterococcus faecalis* bacteria and fungi, but it could not dissolve tissue. Gambier extract (*UncariagambirRoxb*) has antioxidant and antibacterial with phenolic content in the form of catechins. **Purpose:** Describe the antibacterial comparison of gambier extract (*Uncaria gambir Roxb*) and 2% chlorhexidine against *Enterococcus faecalis* bacteria. **Material and Methods:** This research is a laboratory experimental research. *Enterococcus faecalis* ATCC 29212 is a microbiological sample. Extract of gambier (*UncariagambirRoxb*) with concentrations of 1%, 1.5%, 2%, 2.5%, and 3%. 2% chlorhexidine solution and a paper disc to measure the inhibition zone were placed on a petri dish containing Mueller-Hinton agar medium, and incubated 1x24 hours. Gambier extract was made by maceration and rotavapor methods. **Results:** Gambier extract (*UncariagambirRoxb*) and 2% chlorhexidine affected the growth inhibition of *Enterococcus faecalis* bacteria by forming a clear zone around the paper disc in each petri dish. Concentration of 3% is the largest value of the inhibition zone formed in the extract of gambeir (*UncariagambirRoxb*), while chlorhexidine 2% shows the largest inhibition zone, this can be seen from the average inhibition zone formed of 17.20 mm. **Conclusion:** There are differences in the antibacterial of gambier extract (*UncariagambirRoxb*) and 2% chlorhexidine against *Enterococcus faecalis* bacteria.

Keywords: *Enterococcus faecalis*, 2% chlorhexidine, gambier extract (*UncariagambirRoxb*).

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INTRODUCTION

Endodontic treatment is the treatment of pulp disease by removing bacteria and their metabolic products from the root canal system. The purpose of endodontic treatment are to clean and disinfect the root canal system so it can reduce bacteria, remove necrotic tissue, and assist the periapical healing process.¹ Endodontic treatment can be divided into three main stages: access, preparation, and obturation.²

One of the steps of biomechanical preparation in endodontic treatment is root canal irrigation which aims to eliminate bacteria in the root canal. Root canal irrigation has two purposes, mechanical and biological. The mechanical purpose is to remove debris, lubricate the root canal and remove organic and inorganic tissue while the biological purpose is as an antimicrobial.³

To choice of irrigation solution need a good knowledge and understanding of the properties of various irrigation solutions, appropriate irrigation methods and knowledge of various microorganisms that use in the process of root canal infection which contributes to the effectiveness of irrigation solutions. In addition, the effectiveness of the irrigating agent depends on the amount of irrigation solution, the diameter of the root canal, and the condition of the pulp.⁴

Infections that occur in the root canal are caused by microorganisms that gain access to the pulp tissue and periapical. In the case of failure of endodontic treatment, *Enterococcus faecalis*(*E. faecalis*) is one of the most bacteria found in root canals. *E. faecalis* found up to 77% of endodontic treatment failure cases. These bacteria are difficult to remove even though chemo mechanical instrumentation and root canal medication have been performed. Clinical features due to the virulence of this bacterium are acute apical periodontitis, chronic periodontitis, exacerbation of apical periodontitis, marginal periodontitis and periradicular abscess.⁵

Materials that can be used as root canal irrigation solutions are sodium hypochlorite solution, chelator solution or ethylene diamine tetraacetic acid (EDTA), mixture of tetracycline, anacid and a detergent (MTAD), chlorhexidine, and potassium iodide iodine (IPI). Irrigation materials that are commonly used are those that have antiseptic properties and materials that can inhibit the growth of microorganisms in vitro and in vivo in living tissue.^{6,7}

Chlorhexidine is one of the solutions used as root canal irrigation. Chlorhexidine has been used in various concentrations of 0.1-2%. Chlorhexidine 2% is recommended as a root canal irrigant because non-toxic, reduces and eliminates microorganisms from the root canal. Chlorhexidine is used as a disinfectant because it has good antimicrobial properties against gram-positive, gram-negative bacteria, bacterial spores, lipophilic viruses, fungi and dermatophytes. Chlorhexidine 0.1%-0.2% is an antiseptic that is generally used to control oral plaque. Chlorhexidine cannot be used as a sole irrigation solution in root canal treatment because it does not have the ability to dissolve necrotic tissue.⁷

In Indonesia, there are various types of plants that can be used as medicinal plants, one of which is the gambier plant (*Uncaria gambir Roxb*). Gambier (*Uncaria gambir Roxb*) comes from Sumatra and Kalimantan. Gambier is a plant with a height of 1-3 cm.⁸ Gambier has been used as a complement to chewed betel and is believed to strengthen teeth. The main content of Gambier extract is about 7-33% catechins, a polyphenolic compound that has the potential as an antioxidant and antibacterial.⁹

Based on several studies on gambir, Hafsa (2016) researched the benefit of antibacterial gambier and obtained the results that Gambier extract had antibacterial activity against *Enterococcus faecalis* bacteria with a minimal inhibitory concentration of 1%.¹⁰ A similar study conducted by Merta (2013) resulted in the concentration of the inhibitory capacity of gambier extract being 80-90%

effective against the growth of *Staphlococcus aureus*.¹¹ Krisnawati's research (2009) stated that the combination of concentration and contact time affects the antibacterial effect of gambier extract.¹²

METHODS

The type of research is experimental laboratory research. This research was conducted at the Bandung Institute of Technology Laboratory. Bacteria *Enterococcus faecalis* ATCC 29212 microbiological samples in all experimental stages have been cultured. Extract of gambier (*Uncaria gambir Roxb*) with concentrations of 1%, 1.5%, 2%, 2.5%, and 3% and 2% chlorhexidine solution. Each study group used 6 samples, therefore for the 3 groups required 18 samples.

Gambier extract was made using maceration and rotavapor methods.¹³ Gambier weighed as much as 500 g, then the gambier put into a round bottom flask and 1 L of ethanol was added, after that it stored for 48 hours. During storage, stir 2 times in 1 day.¹⁴

The macerated gambier was filtered and then evaporated at a temperature of 60 C for \pm 1 hour until a thick extract was obtained. After that, let it free for 2 hours so that the extract dries. The gambier extract was weighed using an analytical balance, each weighing 0.1 g, 0.15 g, 0.2 g, 0.25 g, 0.3 g, which was obtained from previous studies.^{15,16,17}

The gambier extract dissolved with 2 ml of 10% DMSO solution to obtain a concentration of 1%, 1.5%, 2%, 2.5%, 3%. After that, the concentration of gambier extract was put into a vial bottle and labeled according to its concentration.

BHIB media (37 gr/1 liter or 3.7gr/100ml) in a closed tube was sterilized by autoclaving at 121 C for 15 minutes. Then pure bacteria *Enterococcus faecalis* which in the reaction tube was inserted into the BHIB media using a round loop. Then incubated at 37°C for 24 hours. After that, the bacteria were put back into sodium agar in a petri dish using a round loop with three

quadrants etched and then incubated at 37°C for 24 hours to see if any bacterial colonies were formed.

Eighteen petri dishes containing Mueller Hinton Agar (MHA) media were prepared. A cotton swab was dipped into a reaction tube which containing *Enterococcus faecalis* bacteria. The cotton swab was scratched on the surface so that the MHA medium was in a petri dish and spread evenly. The next step, the paper disc was placed on a filled petri dish with MHA agar and incubated for each concentration of gambier extract and 2% chlorhexidine using a micropipette. Replication was made three times.¹⁵

Petri dishes were incubated at 37°C for 1x24 hours and after 24 hours the zone of inhibition was measured using a caliper.

RESULT

The results of the research on the measurement of the inhibition zone can be seen in the figure and table as follows:

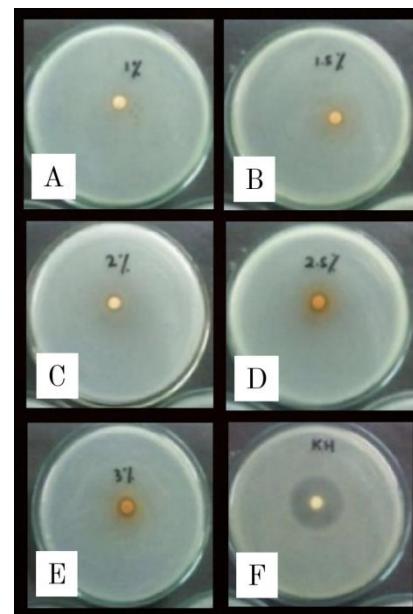
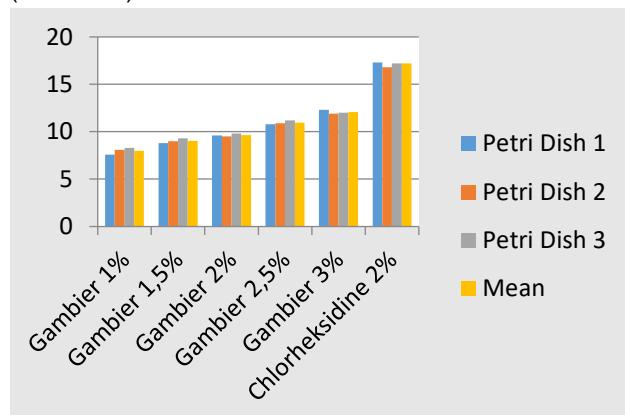


Fig 1. Comparison test results of gambier extract (*Uncaria gambir Roxb*) and 2% chlorhexidine against *Enterococcus faecalis* bacteria after incubation for 24 hours at 37 C. (A) 1% gambier extract (B) 1.5% gambier extract (C) gambier extract 2 % (D) 2.5% gambier extract (E) 3% gambier extract (F) 2% chlorhexidine.

In Figure 1 gambier extract (*UncariagambirRoxb*) and 2% chlorhexidine affect the growth inhibition of *Enterococcus faecalis* bacteria which can be seen in the clear zone around the paper disc in each petri dish.

Chart 1. Measurement Results Inhibition Mean (millimeter)



Based on chart 1, it can be seen that the average diameter of the inhibition zone formed around the paper disc extract of gambier (*UncariagambirRoxb*) concentration of 3% is the largest value compared to the inhibition zone formed at concentrations of 1%, 1.5%, 2%, 2.5%, while chlorhexidine 2% showed the largest inhibition zone, this can be seen from the average inhibition zone formed of 17.20 mm.

Data normality test uses Shapiro-Wilk because the research sample is less than 50 data. Data normality test determines whether the data collected from 18 samples is normally distributed or not, the data distribution is said normal if $p>0.05$. The results of normality test antibacterial comparison of gambier extract (*UncariagambirRoxb*) and 2% chlorhexidine against *Enterococcus faecalis* bacteria showed that all samples studied had $Sig.$ values >0.05 ($p>0.05$) then the data distribution is normal, continued to test variance.

The results of Anova test shows there is a significant difference in antibacterial power from the results of the inhibition zone measurements of various concentrations of gambier extract (*UncariagambirRoxb*) with 2% chlorhexidine against *Enterococcus faecalis* bacteria. This is showed by the value of $Sig.$ 0.000 ($p<0.05$). One Way Anova test can

only show whether or not there is a difference in antibacterial power between gambier extract (*UncariagambirRoxb*) and 2% chlorhexidine against *Enterococcus faecalis* bacteria. For this reason, it is necessary to test using the Multiple Comparison LSD (Least Significant Difference) test in order to know how big the difference in the antibacterial power of each group.

The results of Multiple Comparison (LSD) test show there is a significant difference in the average comparison of the inhibition zones between one group and another. This is showed by the value of $p<0.05$ in all comparisons between various concentrations of gambier extract (*UncariagambirRoxb*) and 2% chlorhexidine.

DISCUSSION

Enterococcus faecalis has been known as a bacterium that can survive in endodontic treatment. The frequency of the presence of *E. faecalis* in cases of post-endodontic infection was found to be nine times higher than that of primary endodontic infections.^{3,4,5}

The ability of a bacterium to survive against medicine can be known by looking at the decreasing colonies of a bacterium and the concentration of medicine liquid is one of the factors that affect the effectiveness of the medicine in killing *E faecalis*.^{18,19}

The results are each concentration of gambier extract there was an inhibition zone against *E faecalis* bacteria. In the 2% chlorhexidine test,²⁰ the inhibition zone against the bacteria *E. faecalis* formed had a larger size than the inhibition zone in the gambier extract. The size of the inhibition zone in 2% chlorhexidine on average is 17.20 mm, while the gambier extract with a concentration of 1% has an inhibition zone of 8.00 mm, a concentration of 1.5% with an inhibition zone of 9.03 mm, a concentration of 2% with an inhibition zone of 9.63 mm, a concentration of 2.5% with an inhibition zone of 10.97 mm and a concentration of 3% having an inhibition zone of 12.07 mm.

The inhibition zone formed after an incubation period of 24 hours indicates that the bacteria in the area cannot grow due to the influence of the test material, which is gambier extract.²¹ In this study, it is shown that the concentration of gambier extract which increased from 1%, 1.5%, 2%, 2.5% and 3% to *E. faecalis* bacteria for 24 hours showed a significant increase in the inhibition zone ($p<0.05$) much bigger the concentration of the gambier extract solution, much bigger the inhibition zone formed. This research is in line with the research by Hafsa et al. showed that the concentration of 1% gambier extract with a contact time of 24 hours against *Enterococcus faecalis* bacteria effectively inhibited the growth of *Enterococcus faecalis* bacteria.¹⁰

Research by Istiyaningsih (2012), also used gambier extract (UncariagambirRoxb) against root canal microbes in necrotic teeth with various concentrations, found that gambier extract was also effective in inhibiting the growth of *Streptococcus mutans* bacteria.²²

Research by Kresnawaty and Zainuddin (2009) stated that methyl derivatives of ethanol extract of gambier leaves has antioxidant and antibacterial activity.^{12,18}

Chlorhexidine is an irrigation solution that has been proven effective against *Enterococcus faecalis* bacteria. Darmetto et al (2005) assessed that the antimicrobial power of 2% chlorhexidine against *E. faecalis* bacteria was more effective than NaOCl, because 2% chlorhexidine is effective as a root canal disinfection agent because its antibacterial capacity can destroy the integrity of bacterial cell membranes, which is changes in cytoplasmic membrane permeability can increase cytoplasmic protein deposition, change cellular osmotic balance, interfere with metabolism, growth and division of bacterial cells, so that *Enterococcus faecalis* cell walls can be damaged, lysed and eventually die.^{19,23}

Based on the results of the research, it was found that gambier extract (UncariagambirRoxb) and 2% chlorhexidine had antibacterial ability in inhibiting the growth of

Enterococcus faecalis ATCC 29212 bacteria. Gambir extract with the smallest concentration of 1% still could inhibit the growth of *Enterococcus faecalis* ATCC 29212 bacteria. This is indicated by the presence of an inhibition zone formed. Thus gambier extract can inhibit the growth of *Enterococcus faecalis* ATCC 29212 bacteria, but 2% chlorhexidine has greater antibacterial power.²⁰

CONCLUSION

There is a difference in the antibacterial of gambier extract (UncariagambirRoxb) and 2% chlorhexidine against *Enterococcus faecalis* bacteria. Chlorhexidine 2% has a greater zone of inhibition against *Enterococcus faecalis* bacteria than gambier extract (UncariagambirRoxb).

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